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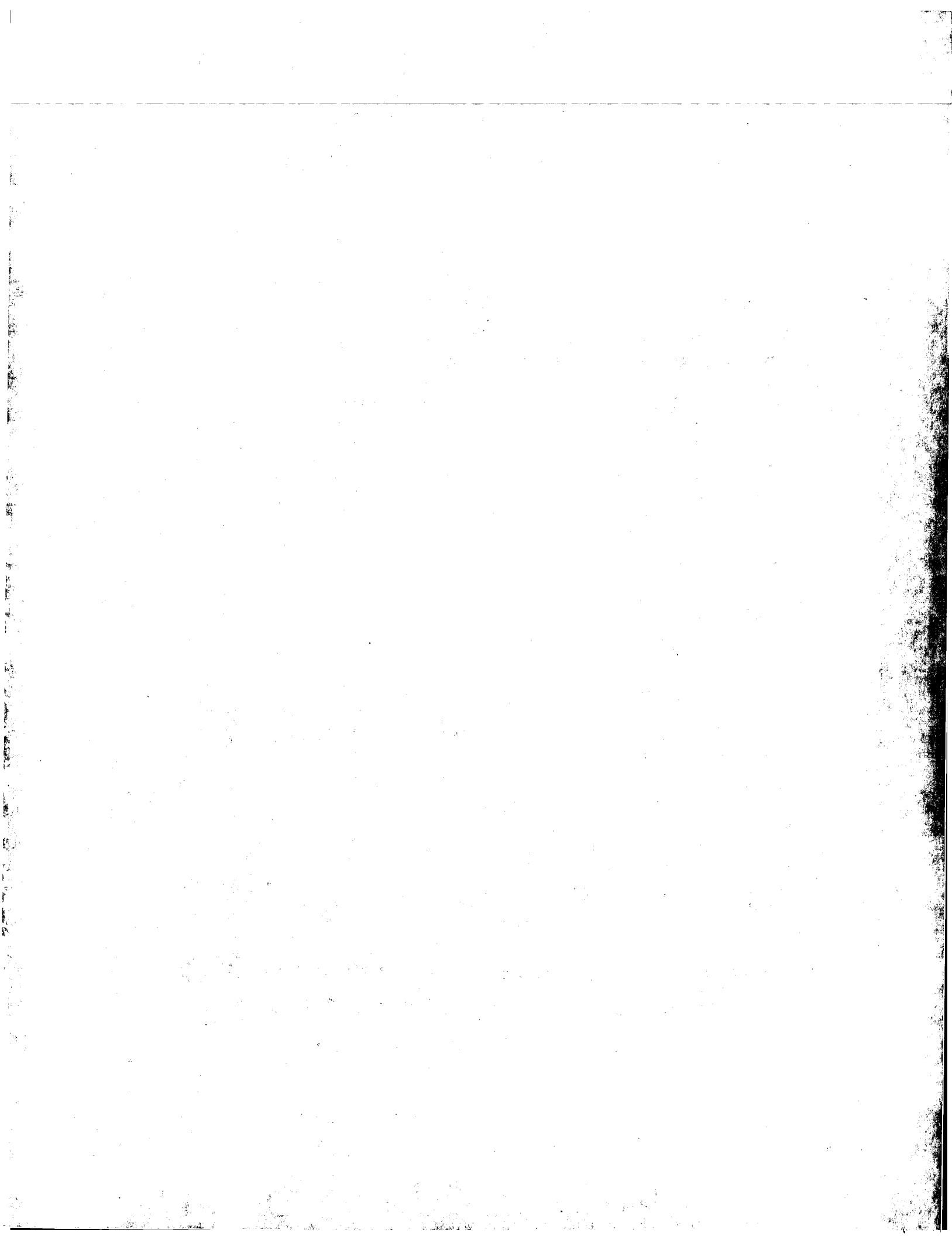
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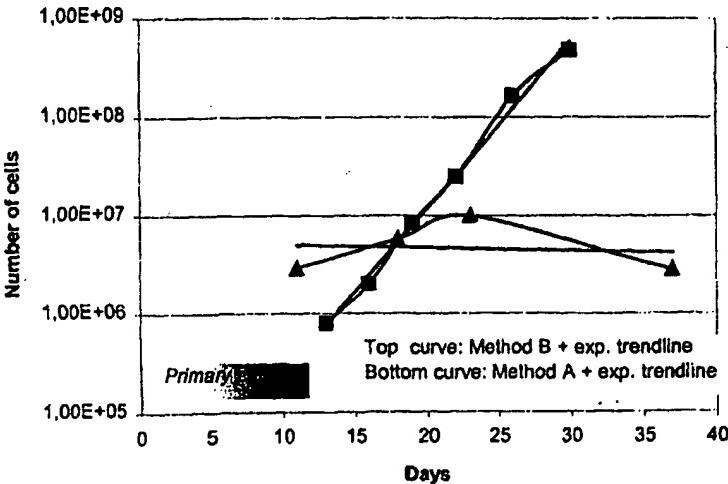
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(54) Title: CELL CULTURE MEDIUM CONTAINING GROWTH FACTORS AND L-GLUTAMINE

Comparison of growth curves of HBMC's cultured according to method A and B.



(57) Abstract: The invention relates to a culture medium for culturing cells, in particular human cells in a process for tissue engineering bone. The medium comprises glucose, a mineral, a vitamin, a growth factor and L-glutamine, wherein the L-glutamine is present in a concentration of at least 300 mg/L.

WO 01/48147 A1

## CELL CULTURE MEDIUM CONTAINING GROWTH FACTORS AND L-GLUTAMINE

The invention relates to the field of tissue engineering. In particular, the invention discloses a medium and a method for culturing cells

5 The need for replacement parts for the human body, in combination with the shortage of donor tissue and/or organs has lead to the production of replacement tissue by seeding cells onto or into a scaffold. Eventually, this should lead to tissue engineered products ready to be implanted to take over the function of missing or injured body parts.

10 The scaffold defines the construct shape and dimensions of the replacement to be implanted. Preferably, it is manufactured of a porous or fibrous biodegradable material, so that the degradation of the scaffold proceeds parallel with accumulation of tissue components (growth and 15 synthesis of extracellular matrix (ECM)). Thus, the function of the scaffold, the provision of shape and strength, will gradually be taken over by the formed tissue components.

In view of the fact that cells from allogenic sources are generally rejected, autologous cells isolated from a 20 tissue biopsy from the patient to be treated are preferably used. In order to minimize the size of the biopsy needed and to minimize the time required for cell expansion, the expanded cells have to be first applied in/onto the scaffold in an efficient manner. In addition, the cells should be 25 distributed homogeneously throughout the scaffold, in order to enable continuous neo-tissue formation.

In general, the cells that have been harvested from the patient's body are cultured *in vitro* for a certain period of time, either with or without a scaffold material present. 30 During this culturing period, proliferation and/or differentiation of the cells may take place, depending on the type of cells harvested and on the objective type of tissue.

In the literature and on the market, various cell culture media are known. These usually contain glucose, 35 inorganic salts (minerals), amino acids, and vitamins. Other

ingredients that are sometimes used include ribonucleosides, deoxyribonucleosides and antibiotics. Well-known, commercially available culture media are for instance Dulbecco's Modified Minimal Eagle's Medium (DMEM) and Alpha 5. Minimal Eagle's Medium ( $\alpha$ -MEM).

In the US patent 5,197,985, a method is disclosed for enhancing the implantation and differentiation of marrow-derived mesenchymal cells. The method is stated to be particularly intended as a means for treating skeletal and 10 other connective tissue disorders in humans. For the culturing of the mesenchymal cells, a medium was employed that comprised the commercially available BGJ<sub>b</sub> medium (Fitton-Jackson modification) and selected lots of 10% fetal bovine serum. Further, the medium F-12 Nutrient Mixture (Ham) 15 was used for selective marrow-derived mesenchymal cell separation.

The production of tissue engineered products will generally only commence once the type of injury or disorder is known and a specific treatment has been decided upon. 20 While the tissue engineering is carried out, the patient is in the meantime suffering from his injuries or disorder. Thus, in order to minimize a patient's discomfort it is of great importance that the production of engineered tissues proceeds as fast as possible. A disadvantage of most of the 25 known culture media is that they do not allow for a sufficiently fast proliferation and/or differentiation of the cells which are cultured in it. This disadvantage is particularly apparent in case human cells are cultured.

The present seeks to provide a culture medium wherein 30 cells proliferate and/or differentiate very fast. The objective culture medium is to allow for a reduction of the time needed for culturing cells in the treatment of a patient via a tissue engineering approach, when compared to conventional culture media.

35 Surprisingly, it has now been found that the rate of proliferation and/or differentiation of cells can be

significantly increased by culturing them in a medium which comprises a growth factor and an increased concentration of L-glutamine. The invention accordingly relates to a culture medium comprising glucose, a mineral, a vitamin, a growth factor and L-glutamine, wherein the L-glutamine is present in a concentration of at least 300 mg/L.

Human cells have been found to proliferate much faster in a culture medium according to the invention than in conventional culture media. Moreover, the differentiation of cells, which are capable of differentiation, has also been found to proceed faster than in conventional culture media. This increased cultivation rate is advantageous when the culture medium is employed for manufacturing a tissue engineered product for treatment of a patient.

The culture medium comprises several components, which are dissolved or suspended in a suitable liquid, preferably distilled water. As cells will be cultured in the medium, it will be clear that it is of advantage to work under sterile conditions to prevent microbial contamination of the cultures.

In order for cells to be cultivated, it is necessary that sufficient nutrients are present in the culture medium. To this end, the present culture medium comprises glucose, minerals and vitamins.

Glucose is an important nutrient in a culture medium. In accordance with the invention it is preferred that the concentration of glucose is at least 750 mg/L, more preferred between 900 and 2000 mg/L.

The minerals may suitably be chosen from the group of calcium, potassium, lithium, magnesium, sodium, sulphate, chloride, bicarbonate, dihydrogenphosphate ions, and combinations thereof. Particularly suitable minerals are calcium chloride, preferably in an amount of 100-400 mg/L, potassium chloride, preferably in an amount of 200-600 mg/L, sodium chloride, preferably in an amount of 5500-8000 mg/L, magnesium sulphate, preferably in an amount of 100-300 mg/L,

sodium bicarbonate, preferably in an amount of 1500-3000 mg/L, and sodium dihydrogenphosphate, preferably in an amount of 100-200 mg/L. The concentrations of these minerals may be varied within rather wide ranges. Typically, the combined 5 amount of minerals in a culture medium according to the invention is chosen such as to result in near physiological conditions (around 0.9%).

The vitamins may suitably be chosen from the group of L-ascorbic acid, biotin, D-calcium pantothenate, choline 10 chloride, folic acid, i-inositol, nicotinamide, pyridoxal, riboflavin, thiamine, vitamin B<sub>12</sub>, vitamin A (retinoic acid) and combinations thereof. These may be present in the following, preferred concentrations:

15	L-ascorbic acid	75 - 1500 mg/L;
	biotin	0.03 - 0.5 mg/L;
	D-calcium pantothenate	0.5 - 2 mg/L;
	choline chloride	0.5 - 2 mg/L;
	folic acid	0.5 - 2 mg/L;
	i-inositol	1 - 4 mg/L;
20	nicotinamide	0.5 - 2 mg/L;
	pyridoxal	0.5 - 2 mg/L;
	riboflavin	0.03 - 0.5 mg/L;
	thiamine	0.05 - 2 mg/L; and
	vitamin B <sub>12</sub>	1 - 2 mg/L.

25 It is further preferred that the present culture medium comprises amino acids. Suitably, these amino acids are selected from the following group in the specified concentrations:

30	L-alanine	10 - 50 mg/L;
	L-arginine	75 - 150 mg/L;
	L-asparagine	30 - 80 mg/L;
	L-aspartic acid	10 - 50 mg/L;
	L-cystine	10 - 50 mg/L;
	L-cysteine	75 - 130 mg/L;
35	L-glutamic acid	50 - 100 mg/L;
	L-alanyl-L-glutamine	5 - 20 mg/L;

	glycine	30 - 80 mg/L;
	L-histidine	20 - 60 mg/L;
	L-isoleucine	30 - 80 mg/L;
	L-leucine	30 - 80 mg/L;
5	L-lysine	50 - 100 mg/L;
	L-methionine	5 - 30 mg/L;
	L-phenylalanine	10 - 50 mg/L;
	L-proline	20 - 60 mg/L;
	L-serine	10 - 50 mg/L;
10	L-threonine	30 - 80 mg/L;
	L-tryptophan	5 - 15 mg/L;
	L-tyrosine	10 - 60 mg/L; and
	L-valine	30 - 80 mg/L.

The combined concentration of the amino acids  
 15 preferably lies between 800 and 5000 mg/L. It is to be noted  
 that this concentration range does not include L-glutamine.

L-glutamine is an important component of a culture  
 medium according to the invention. It has been found that a  
 concentration L-glutamine of at least 300 mg/L, preferably  
 20 between 400 and 800 mg/L leads to a particular high  
 proliferation and/or differentiation rate of cells cultured  
 in the present culture medium.

Another component of a culture medium according to  
 the invention is a growth factor. The nature of the growth  
 25 factor may suitably be chosen dependent on the type of cells  
 to be cultured. Examples of preferred growth factors are Bone  
 Morphogenetic Protein (BMP), Epidermal Growth Factor (EGF),  
 basic Fibroblast Growth Factor (bFGF), Nerve Growth Factor  
 (NGF), Bone Derived Growth Factor (BDGF), Transforming Growth  
 30 Factor-β1 (TGF-β1), and human Growth Hormone (hGH). In a  
 preferred embodiment, the culture medium is employed to  
 culture cells which are used in tissue engineering bone. In  
 accordance with this embodiment, the growth factor is  
 preferably bFGF. The growth factor is preferably present in  
 35 an amount between 0.1 µg/L and 10 µg/L.

In a preferred embodiment, the present culture medium further comprises an antibiotic or a combination of antibiotics. Examples of suitable antibiotics include penicillin G, gentamicin, fungizone, streptomycin. It has surprisingly been found that the nature of the antibiotic influences that cultivation rate of cells in the present culture medium. A highly preferred combination of antibiotics is penicillin G and streptomycin. These antibiotics are preferably each employed in an amount of 50-150 µg/mL, more preferably they are both employed in an amount of 100 µg/mL. The total amount of antibiotics in a culture medium according to the invention preferably lies between 75 and 300 µg/mL.

The culture medium may further comprise any ingredient conventionally employed in culture media. Examples of such ingredients include serum, such as fetal bovine serum, autologous serum or synthetic serum (e.g. Ultrocer®), thioctic acid, phenol red, sodium pyruvate, ribonucleosides and deoxyribonucleosides. A ribonucleoside or a deoxyribonucleoside is preferably present in a concentration of between 5 and 15 mg/L. It is to be noted that when serum is used, it is added after the formulation of the culture medium. In other words, the amount of serum added dilutes the concentrations mentioned herein. Fetal bovine serum may be added in such an amount that the composition of serum and culture medium comprises between 5 and 15 vol.% of serum. Ultrocer® may be added in such an amount that the composition of serum and culture medium comprises between 1 and 10 vol.% of serum.

As has been mentioned above, the present culture medium is particularly useful for culturing human cells, e.g. in a process for manufacturing tissue engineered products, such as skin grafts, bone implants, cartilage implants. The present culture medium has particularly been found advantageous for the *in vitro* production of bone tissue. More osteogenic cells are formed resulting in a higher success of bone formation after implantation, when the cells are

cultured in a culture medium in accordance with the invention when compared to conventional culture media.

In principle, cells of any type may be cultivated in a culture medium according to the invention. Preferred cell types include stem cells, progenitor cells, mesenchymal cells, epithelial cells, cartilaginous cells, osseous cells, muscular cells, gland cells, fat cells, pericytes, satellite cells and dermal cells. Highly preferred cells to be cultured in the present culture medium are progenitor cells which may differentiate into bone. Differentiation of the cells may be facilitated by the presence of growth factors, such as Bone Morphogenetic Protein, or dexamethasone, which is preferably used in an amount of between  $1 \times 10^{-9}$  and  $1 \times 10^{-7}$  µg/L. The cells may be cultured in the presence of a scaffold or without a scaffold. If the cells are cultured in the presence of a scaffold, they can first suitably be seeded onto or into the scaffold in any known manner.

The invention will now be elucidated by the following, non-restrictive examples.

20

Example I

A culture medium was prepared by admixing the following ingredients in the specified concentrations in water:

25

	Component	1x Liquid mg/L
<b>INORGANIC SALTS</b>	CaCl <sub>2</sub> . 2H <sub>2</sub> O	264.00
	KCl	400.00
	MgSO <sub>4</sub> . 7H <sub>2</sub> O	200.00
	NaCl	6800.00
	NaHCO <sub>3</sub>	2200.00
	NaH <sub>2</sub> PO <sub>4</sub> . 2H <sub>2</sub> O	158.00
<b>OTHER COMPONENTS</b>	D-Glucose	1000.00
	DL-68 Thioctic Acid	0.20
	Phenol Red	10.00
<b>AMINO ACIDS</b>	Sodium Pyruvate	110.00
	L-Alanine	25.00
	L-Arginine . HCl	127.00
	L-Asparagine . H <sub>2</sub> O	50.00

	L-Aspartic Acid	30.00
	L-Cystine	24.00
	L-Cysteine HCl	100.00
	L-Glutamic Acid	75.00
	Glycine	50.00
	L-Histidine HCl . H <sub>2</sub> O	42.00
	L-Isoleucine	53.00
	L-Leucine	52.00
	L-Lysine . HCl	73.00
	L-Methionine	15.00
	L-Phenylalanine	32.00
	L-Proline	40.00
	L-Serine	25.00
	L-Threonine	48.00
	L-Tryptophan	10.00
	L-Tyrosine	36.00
	L-Valine	46.00
VITAMINS	L-Ascorbic Acid	50.00
	Biotin	0.10
	D-Ca Pantothenate	1.00
	Choline Chloride	1.00
	Folic Acid	1.00
	l-Inositol	2.00
	Nicotinamide	1.00
	Pyridoxal HCl	1.00
	Riboflavin	0.10
	Thiamine HCl	1.00
	Vitamin B <sub>12</sub>	1.40
RIBONUCLEOSIDES	Adenosine	10.00
	Cytidine	10.00
	Guanosine	10.00
	Uridine	10.00
DEOXYRIBONUCLEOSIDES	2'Deoxyadenosine	10.00
	2'Deoxycytidine HCl	11.00
	2'Deoxyguanosine	10.00
	2'Deoxythymidine	10.00
	penicillin G	100 µg/mL
	streptomycin	100 µg/mL
	bFGF	1 ng/mL
5	L-glutamine	584 mg/L

Example II

Introduction.

In order to culture Human Bone Marrow Cells (HBMC's) efficiently it is necessary to develop a suitable cell culture medium. We compared two different media to investigate which type is most suitable for culturing HBMC's and to see if there is a difference in, *in vivo*, bone

formation after culturing these cells on a CaP scaffold. We performed several studies on HEMC's obtained of several different patients using either  $\alpha$ -MEM containing 10% Fetal Bovine Serum (FBS), 0.2mM L-ascorbic acid 2-phosphate (AsAP), 5 0.1mg/ml penicillin G, 50 $\mu$ g/ml gentamicin and 0.3 $\mu$ g/ml fungizone (AB) or in  $\alpha$ -MEM containing 10% FBS, 0.2mM L- ascorbic acid 2-phosphate (AsAP), 1ng/ml basic Fibroblast Growth Factor (bFGF), 2mM L-glutamine (L-Glu), 100U/ml penicillin, 100 $\mu$ g/ml streptomycin (pen/strep) and 1% (50U/ml) 10 heparin (added in primary cultures only) and compared morphology, growth rates and bone formation. It was found that the combination of L-ascorbic acid, heparin, bFGF, L-Glutamine and penicillin/streptomycin in our culture medium enhanced cell-growth and showed a higher extent of bone 15 formation.

#### Materials and Methods.

##### Human Bone Marrow cell collection and culture

20

###### Method A

Bone marrow aspirate was obtained from the iliac crest of a patient. In short 5 ml of aspirate was resuspended in 20 ml  $\alpha$ -MEM containing 50U/ml heparin and 10% fetal bovine 25 serum. The cell suspension is then resuspended using a 20G needle and centrifuged for 10 minutes at 300 g. The supernatant is discarded and the cell pellet is resuspended in  $\alpha$ -MEM containing 10%FBS, 0.2mM L-ascorbic acid 2-phosphate (AsAP), 0.1mg/ml penicillin G, 50 $\mu$ g/ml gentamicin and 0.3 $\mu$  30 g/ml fungizone (AB). The obtained mononucleated cells are then plated at a density of  $\pm$ 500.000 cells/cm<sup>2</sup> in tissue culture flasks. Cells were grown at 37°C with 5% CO<sub>2</sub> in a humid atmosphere. The culture medium is refreshed twice a week and at near confluence the adherent cells are washed 35 with phosphate buffered saline solution (PBS) and enzymatically released by incubating the cells with a 0.25%

Trypsin-EDTA solution at 37°C for at least 10 Minutes. The released cells are then thoroughly resuspended and replated at a density of 5000-10.000 cells/cm<sup>2</sup> subsequent passages (up to the fifth passage) are performed when cells reach near confluence and cell morphology is monitored with light microscopy.

5  
Method B

Bone marrow aspirate was obtained from the iliac crest of a patient. In short 5 ml of aspirate was resuspended in 20 ml α-MEM containing 50U/ml heparin and 10% fetal bovine serum. The cell suspension is then resuspended using a 20G needle and centrifuged for 10 minutes at 300 g. The supernatant is discarded and the cell pellet is resuspended in α-MEM containing 10%FBS, 0.2mM AsAP, 1ng/ml bFGF, 2mM L-glutamine (L-Glu), 100U/ml penicillin and 100μg/ml streptomycin (pen/strep) and 1% (50U/ml) heparin (added in primary cultures only). The obtained mononucleated cells are then plated at a density of ±500.000 cells/cm<sup>2</sup> in tissue culture flasks. Cells were grown at 37°C with 5% CO<sub>2</sub> in a humid atmosphere. The culture medium is refreshed twice a week and at near confluence the adherent cells are washed with phosphate buffered saline solution (PBS) and enzymatically released by incubating the cells with a 0.25%Trypsin-EDTA solution at 37°C for at least 10 Minutes. The released cells are then thoroughly resuspended and replated at a density of 5000-10.000 cells/cm<sup>2</sup> subsequent passages (up to the fifth passage) are performed when cells reach near confluence and cell morphology is monitored with light microscopy.

30  
In Vivo experiments and Histology.

Porous CaP particles of size 2 by 3 mm are used for culturing the released HBMC's on. In short harvested HBMC's of several passages were seeded in a density of 100.000-200.000 cells/particle. The cells were cultured during one

week using  $\alpha$ -MEM containing 10%FBS, 0.2mM AsAP, 10 nM Dexamethason (Dex) and 10mM beta glycerophosphate ( $\beta$ GDP) before implantation.

Shortly before implantation the samples were soaked  
5 in  $\alpha$ -MEM, washed in PBS and subcutaneously implanted into  
nude mice and kept *in vivo* for 4-6 weeks. Control samples  
incubated in both media, without cells were also implanted.  
At the end of the *in vivo* period the implanted samples were  
removed and immediately fixated in 1.5% glutaraldehyde in  
10 0.14 M cacodylic acid buffer, pH 7.2-7.4.

After dehydration in an alcohol series and embedding  
in methyl methacrylate, the samples are sectioned on a  
Histological diamond innerlock saw (Leyden Microtome cutting  
system). Sections of around 10 $\mu$ m are stained with basic  
15 fuchsin and methylene blue, in order to study bone formation.  
The sections were then scored per patient for bone formation.

#### Results.

##### 20 Morphology.

Medium with AB (Method A): figure 1 shows that often  
larger, flatter cell morphology observed. Cell growth was  
limited, and there was only occasionally bone formation  
observed after subcutaneous implantation.

25 Improved medium (Method B): figure 2 shows that cells  
with spindle shaped and fibroblastic morphology were  
obtained. Rapid cell proliferation was observed. Also, De  
Novo bone formation after subcutaneous implantation was  
widespread (see fig. 3).

30 Figure 4 shows a comparison of the growth curves of  
the cells cultured with methods A and B.

Figure 5 shows a comparison of 22 patients cultured  
with method A and 15 patients cultured with method B, in  
relation to bone formation.

Discussion.

HBMC's cultured according to method B showed a linear culture expansion compared to HBMC's cultured according to method A. The growth rate of the two methods observed show

5 that culturing HBMC's according to method B, results in a higher growth rate than observed for method A. In time culturing HBMC's with  $\alpha$ -MEM containing 10%FBS, 0.2mM AsAP, 0.1mg/ml penicillin G, 50 $\mu$ g/ml gentamicin and 0.3 $\mu$ g/ml fungizone (method A) shows a decrease in cell growth. Use of

10 culture medium comprising of bFGF, L-Glu, pen/strep, AsAP, with the addition of 50U/ml heparin in primary cultures has shown an improved morphology, growth rate and *in vivo* osteogenic character of the cells, after they have been stimulated to osteoprogenitor differentiation with

15 dexamethasone.

Claims

1. Culture medium comprising glucose, a mineral, a vitamin, a growth factor and L-glutamine, wherein the L-glutamine is present in a concentration of at least 300 mg/L.
2. Culture medium according to claim 1, wherein the L-glutamine is present in a concentration between 400 and 800 mg/L.
3. Culture medium according to claim 1 or 2, wherein the growth factor is chosen from the group of Bone Morphogenetic Protein (BMP), Epidermal Growth Factor (EGF), basic Fibroblast Growth Factor (bFGF), Nerve Growth Factor (NGF), Bone Derived Growth Factor (BDGF), Transforming Growth Factor- $\beta$ 1 (TGF- $\beta$ 1), and human Growth Hormone (hGH).
4. Culture medium according to claim 3, wherein the growth factor is bFGF.
5. Culture medium according to claim 4, wherein the bFGF is present in a concentration between 0.1 and 10  $\mu$ g/L.
6. Culture medium according to any of the preceding claims, wherein the mineral comprises one or more ions chosen from the group of calcium, potassium, lithium, magnesium, sodium, sulphate, chloride, bicarbonate, and dihydrogenphosphate ions.
7. Culture medium according to claim 6, wherein the mineral is present in a concentration between 5 and 15 g/L.
8. Culture medium according to any of the preceding claims, wherein the vitamin is chosen from the group of L-ascorbic acid, biotin, D-calcium pantothenate, choline chloride, folic acid, i-inositol, nicotinamide, pyridoxal, riboflavin, thiamine, vitamin B<sub>12</sub>, vitamin A and combinations thereof.
9. Culture medium according to claim 8 comprising L-ascorbic acid in a concentration between 75 and 1500 mg/L.
10. Culture medium according to any of the preceding claims comprising one or more amino acids chosen from the group of L-alanine, L-arginine, L-asparagine, L-aspartic

acid, L-cystine, L-cysteine, L-glutamic acid, L-alanyl-L-glutamine, glycine, L-histidine, L-isoleucine, L-leucine, L-Lysine, L-methionine, L-phenylalanine, L-proline, L-serine, L-threonine, L-tryptophan, L-tyrosine, and L-valine.

5 11. Culture medium according to claim 10, wherein the amino acids are present in a concentration between 0.8 and 5 g/L.

12. Culture medium according to any of the preceding claim comprising an antibiotic.

10 13. Culture medium according to claim 12, wherein the antibiotic is chosen from the group of penicillin G, gentamicin, fungizone, and streptomycin.

14. Culture medium according to claim 13 comprising as antibiotic a combination of penicillin G and streptomycin, 15 wherein both penicillin G and streptomycin are present in an amount of 50-150 µg/mL.

15. Culture medium according to any of the preceding claims comprising distilled water.

16. Culture medium according to any of the preceding 20 claims comprising a ribonucleoside and/or a deoxyribonucleoside.

17. Culture medium according to any of the preceding claims to which serum is added.

18. Use of a culture medium according to any of the 25 preceding claims for culturing human cells.

19. Use according to claim 18, wherein the cells are chosen from the group of stem cells, progenitor cells, mesenchymal cells, epithelial cells, cartilaginous cells, osseous cells, muscular cells, gland cells, fat cells, 30 pericytes, satellite cells and dermal cells.

20. Use according to claim 19, wherein the cells are progenitor cells.



Figure 1

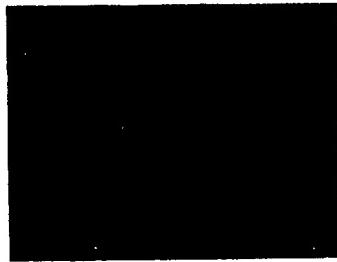


Figure 2



Figure 3

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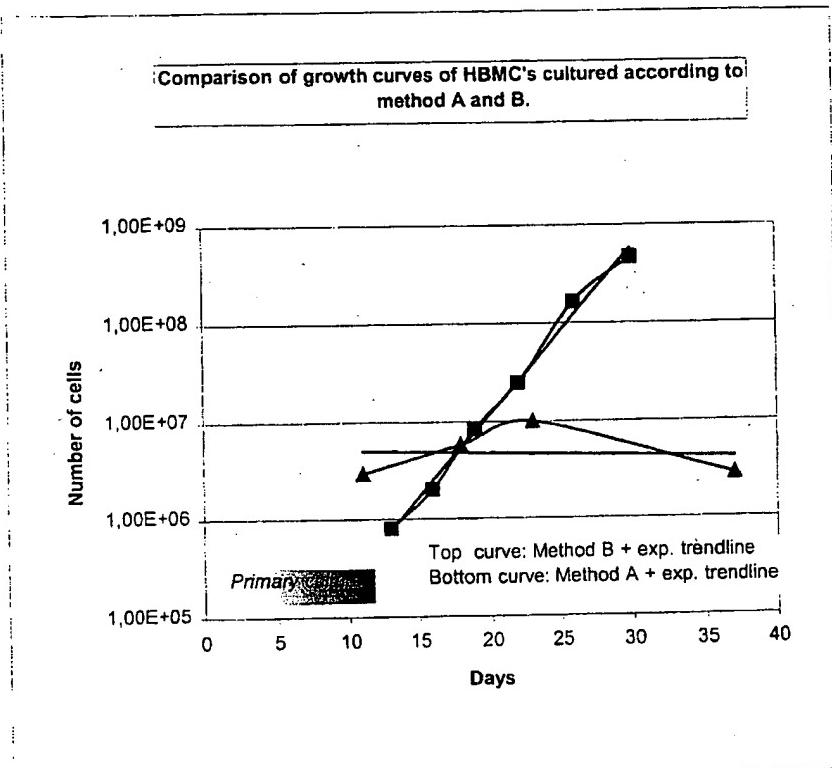


Figure 4

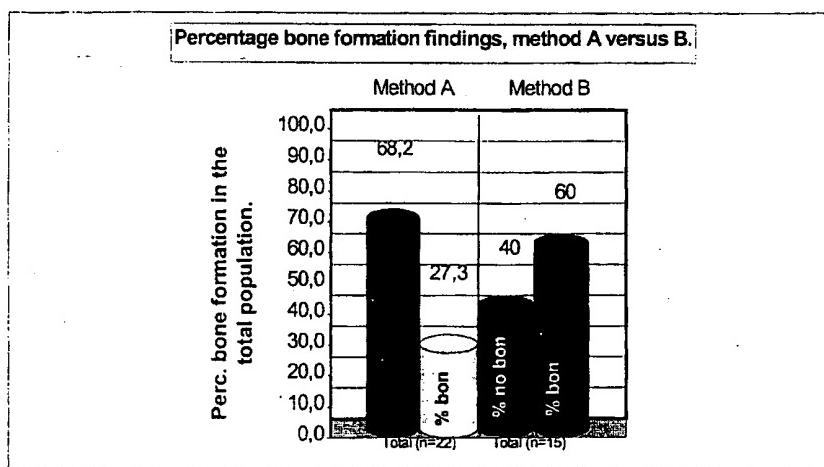


Figure 5

# INTERNATIONAL SEARCH REPORT

International Application No  
PCT/NL 00/00958

**A. CLASSIFICATION OF SUBJECT MATTER**  
IPC 7 C12N5/00

According to International Patent Classification (IPC) or to both national classification and IPC

**B. FIELDS SEARCHED**

Minimum documentation searched (classification system followed by classification symbols)  
IPC 7 C12N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

EPO-Internal, BIOSIS, WPI Data, CHEM ABS Data

**C. DOCUMENTS CONSIDERED TO BE RELEVANT**

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	US 5 830 682 A (MOORE EMMA E) 3 November 1998 (1998-11-03) * column 8, line 37- line 67; column 20, line 20- line 26; column 24, line 41- line 55; Table 1, claim 1 *	1-20
X	US 5 858 783 A (SPAULDING GLENN F ET AL) 12 January 1999 (1999-01-12) * column 7, line 29- line 42; column 8, line 29 - line 39; column 12, line 48 - line 54; Table 1, claim 1 * column 7, line 31 -column 7, line 42; claim 1; table 1 ---	1-20 -/-

Further documents are listed in the continuation of box C.

Patent family members are listed in annex.

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Date of mailing of the international search report

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# INTERNATIONAL SEARCH REPORT

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C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		
Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 98 16630 A (TEXAS A & M UNIVERSITY SYST ;BAZER FULLER W (US); PIEDRAHITA JORGE) 23 April 1998 (1998-04-23) page 9, line 7- line 14; page 14, line 17 - page 15 line 24; page 39, second paragraph; page 45, line 6 - line 12; claims 1-27 * the whole document ---- US 5 811 094 A (HAYNESWORTH STEPHEN E ET AL) 22 September 1998 (1998-09-22) column 8, line 51 -column 9, line 45 ----	1-20
X		1-20

# INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No
PCT/NL 00/00958

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
US 5830682	A 03-11-1998	US 5648219 A US 5683906 A AU 3586595 A CA 2199377 A EP 0804554 A JP 10505498 T WO 9607733 A	15-07-1997 04-11-1997 27-03-1996 14-03-1996 05-11-1997 02-06-1998 14-03-1996
US 5858783	A 12-01-1999	US 5496722 A US 5962324 A US 6117674 A US 5846807 A	05-03-1996 05-10-1999 12-09-2000 08-12-1998
WO 9816630	A 23-04-1998	AU 4984697 A EP 0941310 A	11-05-1998 15-09-1999
US 5811094	A 22-09-1998	US 5486359 A US 5226914 A US 5197985 A US 6010696 A US 6087113 A US 5733542 A US 5837539 A AU 669850 B AU 2252492 A CA 2111845 A EP 0592521 A JP 7500001 T WO 9222584 A	23-01-1996 13-07-1993 30-03-1993 04-01-2000 11-07-2000 31-03-1998 17-11-1998 27-06-1996 12-01-1993 23-12-1992 20-04-1994 05-01-1995 23-12-1992

